

# Neoehrlichiosis: an emerging tick-borne zoonosis caused by *Candidatus Neoehrlichia mikurensis*

Cornelia Silaghi<sup>1</sup> · Relja Beck<sup>2</sup> · José A. Oteo<sup>3</sup> ·  
Martin Pfeffer<sup>4</sup> · Hein Sprong<sup>5</sup>

Received: 10 March 2015 / Accepted: 27 May 2015  
© Springer International Publishing Switzerland 2015

**Abstract** *Candidatus Neoehrlichia mikurensis* is an emerging tick-borne pathogen causing a systemic inflammatory syndrome mostly in persons with underlying hematologic or autoimmune diseases. As it is neither well-known nor well-recognized, it might be misdiagnosed as recurrence of the underlying disease or as an unrelated arteriosclerotic vascular event. The pathogen is transmitted by hard ticks of the genus *Ixodes* and is closely associated with rodents in which transplacental transmission occurs. Transovarial transmission in ticks has not yet been shown. Infection rates vary greatly in ticks and rodents, but the causes for its spatiotemporal variations are largely unknown. This review summarizes the current state of knowledge on the geographical distribution and clinical importance of *Ca. N. mikurensis*. By elucidating the life history traits of this pathogen and determining more accurately its incidence in the human population, a better assessment of its public health relevance can be made. Most urgent research needs are the in vitro-cultivation of the pathogen, the development of specific serological tests, the determination of the full genomic sequence, the routine implementation of molecular diagnosis in diseased patients with a particular panel of underlying diseases, and promoting the knowledge about neoehrlichiosis among general practitioners, hospital physicians and the risk groups such as forest workers or immune-compromised people to raise awareness about this disease that can easily be treated when correctly diagnosed.

---

✉ Cornelia Silaghi  
cornelia.silaghi@uzh.ch

<sup>1</sup> National Center for Vector Entomology, Institute of Parasitology, Vetsuisse Faculty, University of Zurich, Winterthurerstrasse 266a, 8057 Zurich, Switzerland

<sup>2</sup> Department for Bacteriology and Parasitology, Croatian Veterinary Institute, Zagreb, Croatia

<sup>3</sup> Center of Rickettsiosis and Arthropod-borne Diseases, Hospital San Pedro-Center of Biomedical Research of La Rioja, Logroño, Spain

<sup>4</sup> Institute for Animal Hygiene and Veterinary Public Health, University of Leipzig, Leipzig, Germany

<sup>5</sup> Laboratory for Zoonoses and Environmental Microbiology, National Institute of Public Health and Environment (RIVM), Bilthoven, The Netherlands

**Keywords** *Candidatus Neoehrlichia mikurensis* · Anaplasmataceae · *Ixodes ricinus* · *Ixodes persulcatus* · Rodents · *Myodes* spp. · *Apodemus* spp. · Immunodeficiency · Haematological disorders · Thrombosis · Diagnostics · Treatment

## Introduction

Less than a decade ago, the first human case of neoehrlichiosis was diagnosed in a German patient (von Loewenich et al. 2010). Since then, more than a dozen well documented cases have been published in Europe, and also recently cases in China (Li et al. 2012). Neoehrlichiosis can cause systemic inflammatory infections in immune-compromised persons that can be mistaken for the recurrence of an underlying disease or an unrelated arteriosclerotic vascular event. The causative agent, the intracellular bacterium *Candidatus Neoehrlichia mikurensis*, is considered an emerging tick-borne pathogen in Europe where it is transmitted by *Ixodes ricinus*, the most common hard tick species which acts as vector for several zoonotic tick-borne pathogens in Europe (Heyman et al. 2010). Transovarial transmission in the tick vector has neither been observed in epidemiological studies nor experimentally been proven (Burri et al. 2014). Recent evidence points towards *Ca. N. mikurensis* being very closely associated to several rodent species (Burri et al. 2014; Obiegala et al. 2014), and results from xenodiagnostic experiments with ticks have shown that rodents are competent reservoir hosts for *Ca. N. mikurensis* (Burri et al. 2014). Further, a recent study reported observations of transplacental transmission in rodents (Obiegala et al. 2014).

Even though considerable research efforts have been undertaken in recent years to elucidate the natural endemic cycle and geographical distribution of this pathogen, major parts of its life history traits, transmission mechanisms, host associations, and pathogenicity remain largely unknown. In this review, we summarize the current state of knowledge of *Ca. N. mikurensis* in terms of its distribution in ticks and wildlife, the clinical disease it causes, the diagnostic and treatment options, and its public health relevance. Further, we point out urgently needed research efforts for the future.

## History of discovery

All members of the rickettsial family Anaplasmataceae are intracellular alpha-proteobacteria, most of which are transmitted by or infecting different arthropods or other invertebrates. They all have in common that they reside within membrane-enclosed vacuoles inside cells of their eukaryotic hosts (Dumler et al. 2001). Currently, the Anaplasmataceae include the genera *Anaplasma*, *Ehrlichia*, *Aegyptianella*, *Neorickettsia*, and *Wolbachia*, as well as two candidate genera, '*Candidatus Neoehrlichia*' and '*Candidatus Xenohalictis*', the latter detected in abalone (*Haliotis* spp.) (Friedman et al. 2000; Kawahara et al. 2004). Several of them such as *Ehrlichia chaffeensis*, *Ehrlichia canis*, *Anaplasma phagocytophilum* and *Anaplasma marginale* may cause severe diseases in humans and/or animals (Aubry and Geale 2011; Bowman 2011; Dumler et al. 2007).

The history of discovery of *Ca. N. mikurensis* started in 1999 when data on a taxonomically ungrouped *Ehrlichia* DNA were published which had been detected in engorged *I. ricinus* ticks collected from roe deer in The Netherlands. It was first named after Corrie

Schot as the ‘Schotti-variant’ (Schouls et al. 1999). A retrospective investigation of museum-archived *I. ricinus* female ticks collected in Moldova during 1960 indicated that this pathogen has been around for much longer (Movila et al. 2013b).

Similar DNA sequences were detected in *I. ricinus* and *I. persulcatus* ticks from Baltic countries (Alekseev et al. 2001). Additionally, between 1998 and 2001, DNA of an unknown organism was identified in engorged *I. ricinus* ticks collected from asymptomatic patients in Italy. This organism was suggested to be denominated *Ca. Ehrlichia walkeri* sp. nov. (Koutaro et al. 2005). In 2003, DNA sequences of this organism were also detected in *I. ricinus* ticks from Germany, and these investigations were followed by first explorations on possible reservoir hosts (Von Loewenich et al. 2003). Also in 2003, similar DNA sequences to the above mentioned were found in three brown rats (*Rattus norvegicus*) in China and were called the ‘*Rattus*-variant’ (Pan et al. 2003). During 1998, 1999 and 2003, DNA of the same ‘new’ bacterium was detected in 7 out of 15 brown rats from the Japanese island Mikura (Kawahara et al. 2004). This organism was passaged by intraperitoneal injection of a spleen homogenate of the naturally infected rats in laboratory Wister rats, and first information on the ultrastructure of this organism as well as a phylogenetic analysis became available. Electron microscopy photographs presented them as inclusions in the endothelial cell lining of the splenic sinus in one of the Wister rats 2 months after inoculation (Kawahara et al. 2004). At present, the ultrastructural observations by Kawahara et al. (2004) remain the only ones available for this organism that the authors suggested to denominate ‘*Candidatus Neoehrlichia mikurensis*’. This taxonomy currently is still valid because the bacterium has not yet been cultured. The close genetic similarity of the *16S rRNA* and the *groEL* genes places *Ca. N. mikurensis* in the family Anaplasmataceae.

In a more recent investigation, gene sequences from all the above mentioned publications were compared to each other confirming the identity or very close relationship of the *Ehrlichia*-like organisms with the new proposed species *Ca. N. mikurensis* (Jahfari et al. 2012). It forms a separate cluster in the family Anaplasmataceae, together with the North American *Ca. Neoehrlichia lotoris* which had been detected in tick-infested raccoon populations (*Procyon lotor*) and was successfully isolated in the *Ixodes scapularis*-derived ISE6 tick cell line, and was formally described (Munderloh et al. 2007; Yabsley et al. 2008a). Interestingly, the culture was infectious for raccoons in further experiments but it did not produce a detectable infection in laboratory mice, rats or rabbits (Yabsley et al. 2008b). The analysis of several gene loci showed that *Ca. N. lotoris* is closely related to *Ca. N. mikurensis*, but can be differentiated on the basis of unique *16S rRNA* (98.4–98.8 % identity), *groESL* (90.3–90.6 % identity) or *gltA* (84.4 % identity) gene sequences (Munderloh et al. 2007).

## Characteristics of the bacterium

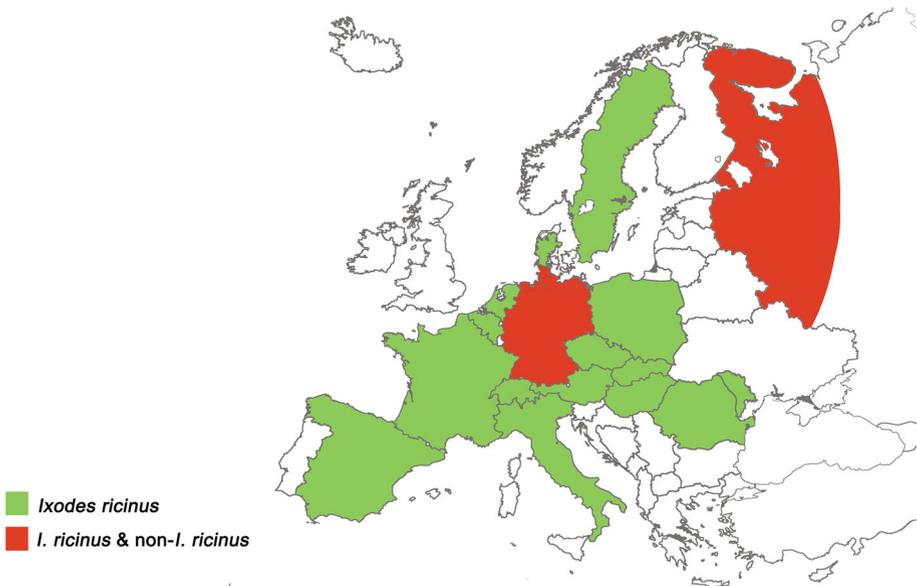
Due to its intracellular character and its presumed endothelial cell tropism, it is highly likely that *Ca. N. mikurensis* will be included in the family Anaplasmataceae, but in a new genus. A full description of *Ca. N. mikurensis* similar to the one recently published for *Ca. N. lotoris* is currently still missing (Yabsley et al. 2008a). Kawahara et al. (2004) described *Ca. N. mikurensis* as a small (0.5–1.5  $\mu\text{m}$ ), gram-negative, pleomorphic coccus of the alpha-proteobacteria which grows in membrane-bound inclusions within the cytoplasm of endothelial cells. *Candidatus N. lotoris* was also described as morphologically similar to

other members of the Anaplasmataceae, but the host cells in raccoons remain elusive. *Candidatus N. lotoris* is also gram-negative, pleomorphic with a size range from 0.5 to 4  $\mu\text{m}$ . The bacterium has a thin periplasmic space that separates the outer unit membrane from the inner cell wall. This is a common characteristic of the family Anaplasmataceae (Yabsley et al. 2008a).

## Ecology of *Candidatus Neohrlichia mikurensis*

### Occurrence and geographical distribution in ticks

*Candidatus N. mikurensis* has so far been detected in *I. ricinus* ticks from 16 European countries covering almost the entire continent (Fig. 1). Overall prevalence rates in *I. ricinus* or *I. persulcatus* ticks collected from the vegetation range from below 1 % to over 20 % (Table 1). Prevalences in *I. ricinus* seem to be somewhat higher than in *I. persulcatus* which usually are around or below 1 %, whereas prevalences in *I. ricinus* seem to range on average around 6–8 %. The prevalence in *I. ricinus* ticks collected from various host animals (including humans) in Europe seems to be actually somewhat lower than in questing ticks, but due to the different study designs, this cannot be quantified (Table 1). Furthermore, *Ca. N. mikurensis* was also detected in other tick species; however, these were usually collected while feeding on host animals (Table 2). Therefore, no evidence is provided thus far that tick species other than *I. ricinus* or *I. persulcatus* are involved in the transmission of *Ca. N. mikurensis*, because its detection in ticks from host animals may be blood meal artefacts. *Candidatus N. mikurensis* has been detected in one questing *Dermacentor reticulatus* tick (Richter et al. 2013), but this number is too low to draw any conclusion from it.



**Fig. 1** *Candidatus Neohrlichia mikurensis* in *Ixodes ricinus* in Europe. *Ixodes* species other than *I. ricinus* include *I. frontalis* in Russia and *I. hexagonus* and *I. trianguliceps* in Germany

**Table 1** Detection of *Candidatus* Neoehrlichia mikurensis in *Ixodes ricinus* ticks in European countries

Country	Tick origin	Number positive/total number (% positive)	Sampling years	References
Austria	Vegetation	22/518 (4.2)	2002–2003	Glatz et al. (2014)
		19/86 (22.1)	na	Derdáková et al. (2014)
Czech Republic	Vegetation	2/20 (10)	na	Richter and Matuschka (2012)
		1473 [in pools] (MIR: 0.4–4.4)	2010	Venclikova et al. (2014)
Denmark	Vegetation	3/138 (2.2)	na	Derdáková et al. (2014)
		3/2625 (MIR: 0.1)	2011	Fertner et al. (2012)
France	Vegetation	2350 [in pools] (MIR: 0.2–0.9)	2008–2012	Michelet et al. (2014)
		1/60 (1.7)	na	Richter and Matuschka (2012)
Germany	Vegetation	2350 [in pools] (MIR: 0.2–1.1)	2008–2012	Michelet et al. (2014)
		189/782 (MIR: 24.2)	2008–2009	Silaghi et al. (2012)
	Vegetation	44/542 (8.1)	na	Richter and Matuschka (2012)
		51/2315 (2.2)	2009–2013	Obiegala et al. (2014)
Rodents		15/234 (6.4)	2010–2011	Silaghi et al. (2012)
		33/965 (3.8)	2012–2013	Obiegala et al. (2014)
Wild boar		1/16 (6.25)	2010–2013	Silaghi et al. (2014)
Humans		9/111 (8.1)	na	Richter and Matuschka (2012)
Dogs		32/773 (4.1)	2010–2011	Kritcken et al. (2013)
Hungary	Vegetation	33/774 (4.3)	2010–2011	Schreiber et al. (2014)
		2004 <sup>a</sup> [9/37 places]	2007	Hornok et al. (2013)
Italy	Vegetation	3/34 (8.8)	2012	Szekeres et al. (2015)
		20/193 (10.5)	2006–2008	Capelli et al. (2012)
Humans		2/433 (0.5)	1995–2011	Orranto et al. (2014)
Vegetation		1/126 (0.8)	1960	Movila et al. (2013b)
Poland	Vegetation	4/1325 (0.3)	2011	Welc-Faleciak et al. (2014a)
Romania	Human	1/1 (100)	2013	Andersson et al. (2014b)

Table 1 continued

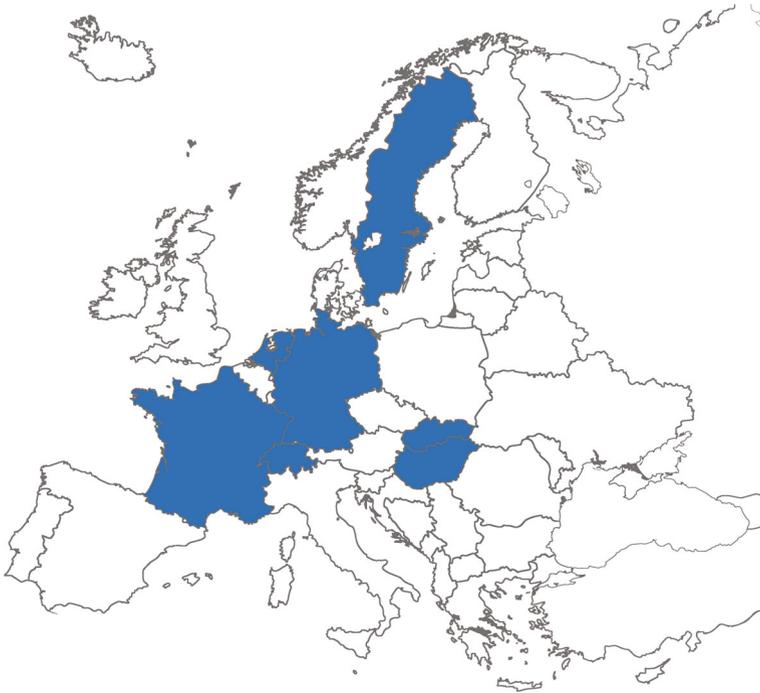
Country	Tick origin	Number positive/total number (% positive)	Sampling years	References
Baltic region	Birds	1/135 (0.7)	2009	Movila et al. (2013a)
Slovakia	Vegetation	2/68 (2.9)	2006	Špitalská et al. (2008)
		16/670 (2.39)	2008–2010	Pangráčová et al. (2013)
		4/378–14/121 (1.1–11.6)	na	Derdáková et al. (2014)
Spain	Cattle	2/200 (1)	2013	Palomar et al. (2014)
Sweden	Vegetation	57/949 (6)	2010–2011	Andersson et al. (2014a)
Switzerland	Vegetation	52/818 (6.4)	2009–2010	Lommano et al. (2012)
	Rodents	1916 (3.5–8)	2009	Maurer et al. (2013)
	Birds	15/575 (2.6)	2011–2012	Burri et al. (2014)
The Netherlands	Vegetation	1205 (3.3)	2007–2010	Lommano et al. (2014)
		300/5343 (5.6)	2000–2009 & 2006–2010	Coipan et al. (2013)
		2350 [in pools] (MIR: 2.4–3.5)	2008–2012	Michelet et al. (2014)
The Netherlands and Belgium	Vegetation, European mouflon, wild boar, sheep	166/2375 (7)	2009–2010	Jahfari et al. (2012)

MIR minimum infection rate, na not available

<sup>a</sup> Number of positives not available

**Table 2** Detection of *Candidatus* Neohhrlichia mikurensis in non-*Ixodes ricinus* tick species

Country	Tick origin	Tick species	Number positive/total number (% positive)	Sampling years	References
Germany	Rodents	<i>Dermacentor reticulatus</i>	9/117 (7.7)	2010–2011	Silaghi et al. (2012)
		<i>Ixodes</i> spp.	2/2 (100)		
		Unidentified larva	1/1 (100)		
	Rodents	<i>Ixodes trianguliceps</i>	1/40	2012–2013	Obiegala et al. (2014)
	Vegetation	<i>D. reticulatus</i>	1/1237 (0.08)	2010–2011	Krücken et al. (2013)
Baltic region Russia (Asiatic part) Russia (Eastern Siberia & Far East) China	Dogs	<i>Ixodes hexagonus</i>	10/151 (6.6)		
	Dogs	<i>I. hexagonus</i>	?/152 (5.9)	2010–2011	Schreiber et al. (2014)
	Birds	<i>Ixodes frontalis</i>	1/4 (25)	2009	Movila et al. (2013a)
	Vegetation	<i>Ixodes persulcatus</i>	2/53 (3.8)	2002	Shpynov et al. (2006)
	Vegetation	<i>I. persulcatus</i>	5/2590 (0.2)	2003–2008	Rar et al. (2010)
	Vegetation	<i>I. persulcatus</i>	8/516 (1.6)	2010	Li et al. (2012)
		<i>Haemaphysalis concinna</i>			
	Vegetation	<i>Ixodes ovatus</i>	?/164 (?)	2000–2001	Kawahara et al. (2004)
	Dogs	<i>Rhipicephalus sanguineus</i> , <i>Haemaphysalis leachi</i>	258 [in pools: 4/76] (5.3)	2011	Kamami et al. (2013)



**Fig. 2** *Candidatus Neohrlichia mikurensis*-infected small mammals as determined by PCR in Europe

### Occurrence and geographical distribution in rodents and other host animals

Similar to other tick-borne diseases, the emergence of human clinical cases corresponds to the distribution of vectors and reservoirs. Studies investigating rodents as well as questing ticks showed that prevalences in rodents were about 2–10 × higher than in ticks (Obiegala et al. 2014; Silaghi et al. 2012). A reservoir function of rodents seems highly likely and was indeed proven in recent xenodiagnostic studies (Burri et al. 2014). Both human cases and naturally infected rodents have been documented in Germany, Sweden and Switzerland whereas so far only infected rodents have been identified elsewhere in Europe (Fig. 2). In Europe, *Ca. N. mikurensis* has been detected in the six rodent species *Apodemus agrarius*, *A. flavicollis*, *A. sylvaticus*, *Myodes glareolus*, *Microtus agrestis* and *Mi. arvalis* (Table 3) and 10 species in Asia (Table 4). Infection rates vary considerably (8.3–52.7 %) among rodent species. *Myodes glareolus* was the most frequently infected species with an average of 9.1 % positive voles. The prevalence in this species varied from 1.8 % in France, up to 52.7 % in Germany. Thus, *My. glareolus* could represent an indicator species in monitoring protocols. Besides *My. glareolus*, other species indicating the presence of *Ca. N. mikurensis* belong to the genera *Apodemus* and *Microtus*. It seems that insectivores do not play a role in the transmission or maintenance of *Ca. N. mikurensis* as so far none of the investigated insectivores were positive in Germany, Sweden and The Netherlands (Andersson and Raberg 2011; Jahfari et al. 2012; Silaghi et al. 2012).

DNA of *Ca. N. mikurensis* has also been detected in *I. ricinus* adults feeding on red deer (*Cervus elaphus*), wild boar (*Sus scrofa*, Silaghi et al. 2014), European mouflon (*Ovis*

**Table 3** Detection of *Candidatus* Neoehrlichia mikurensis in rodents in European countries

Country	Rodent species	Number positive/total number (% positive)	Sampling years	References
France	<i>Myodes glareolus</i>	5/276 (1.8)	2008	Vayssier-Taussat et al. (2012)
Germany	<i>Apodemus flavicollis</i> , <i>A. agrarius</i> , <i>My. glareolus</i>	48/91 (52.7)	2010–2011	Silaghi et al. (2012)
	<i>A. flavicollis</i> , <i>A. agrarius</i> , <i>My. glareolus</i> , <i>Microtus arvalis</i> , <i>Mi. agrestis</i>	36/254 (14.2)	2010–2011	Kricken et al. (2013)
Hungary	<i>My. glareolus</i> , <i>A. sylvaticus</i> , <i>A. flavicollis</i> , <i>Mi. arvalis</i> , <i>Mi. agrestis</i>	181/631 (28.7)	2012–2013	Obiegala et al. (2014)
	<i>A. flavicollis</i>	6/177 (3.4) [in spleen]	2010–2013	Szekeress et al. (2015)
	<i>A. agrarius</i>	6/348 (1.7) [in skin]	2010–2013	Szekeress et al. (2015)
Slovakia	<i>Apodemus</i> spp., <i>My. glareolus</i>	(10.75)	2006–2011	Vichova et al. (2014)
Sweden	<i>My. glareolus</i> , <i>Mi. agrestis</i> , <i>A. sylvaticus</i> , <i>A. flavicollis</i>	68/771 (8.8)	2008	Andersson and Raberg 2011
Switzerland	<i>My. glareolus</i>	50/261 (19)	2008	Andersson et al. (2014a)
	<i>A. sylvaticus</i> , <i>A. flavicollis</i> , <i>My. glareolus</i>	4/103 (3.9)	2011–2012	Burri et al. (2014)
The Netherlands	<i>Apodemus sylvaticus</i> , <i>Mi. arvalis</i> , <i>My. glareolus</i>	11/79 (13.9)	2007–2010	Jahfari et al. (2012)

**Table 4** Detection of *Candidatus Neohelrichia mikurensis* in wild mammals in Asian countries

Country	Rodent species	Number positive/total number (% positive)	Sampling years	References
China	Rodents	8/211 (3.8)	2010	Li et al. (2012)
	<i>Apodemus agrarius</i> , <i>A. peninsulae</i> , <i>Eothenomys custos</i> , <i>Myodes rufocanus</i> , <i>Niviventer confucianus</i> , <i>Rattus norvegicus</i> , <i>Tamias sibiricus</i>	34/841 (4)	2005–2009	Li et al. (2013)
	<i>R. norvegicus</i>	7/15	1998–1999 and 2003	Kawahara et al. (2004)
	Rodents	17/111 (15.3)	2000–2004	Naitou et al. (2006)
	<i>Ap. speciosus</i>	5/55 (9)	2003–2008	Tabara et al. (2007)
Russia (Siberia and Far East)	<i>Ap. argenteus</i>	2/7 (28.6)		
	<i>Microtus</i> spp., <i>A. peninsulae</i> , <i>My. rufocanus</i>	5/1458		Rar et al. (2010)

*orientalis musimon*), sheep (*Ovis aries*) (Jahfari et al. 2012) and cows (Palomar et al. 2014). These findings raised the question on the role of other animals than rodents in the ecology of *Ca. N. mikurensis*. A survey from the Trento region (Italy) revealed *Ca. N. mikurensis* in *My. glareolus* but not in roe deer (*Capreolus capreolus*). Thus, it seems that *Ca. N. mikurensis* cannot infect wild Cervidae (Beninati et al. 2006; Beck et al. 2014b). This assumption was additionally supported by the fact that *Ca. N. mikurensis* has not been found in roe deer (n = 48), red deer (n = 107) and fallow deer (n = 13) in a survey including Slovenia, Croatia and Bosnia and Herzegovina (Beck et al. 2014b). The same study revealed that, in addition to small rodents, *Ca. N. mikurensis*-DNA was present in 30 % of Brown bears (*Ursus arctos arctos*) (3/10), 33 % of European badgers (*Meles meles*) (10/30), 13 % of chamois (*Rupicapra rupicapra*) (7/54) and 1 % of European mouflons (1/62). Interestingly, all 74 investigated red foxes (*Vulpes vulpes*) were negative for *Ca. N. mikurensis*. In a study of northern white-breasted hedgehogs (*Erinaceus roumanicus*) from parks in central Budapest, Hungary (Földvári et al. 2014), *Ca. N. mikurensis*-DNA was present in 2.3 % (2/88) ear tissue samples. Another study from Hungary reported no *Ca. N. mikurensis* detection in birds (Hornok et al. 2014).

## Clinical cases in humans

In Europe, clinical symptoms caused by *Ca. N. mikurensis* infections have mainly been described in immune-compromised patients (Grankvist et al. 2014). Only one case of neehrlichiosis in a previously healthy male patient was thus far reported (von Loewenich et al. 2010). Fourteen cases of neehrlichiosis were reported, with the patients having a median age of 61, being mostly male, and originating from Sweden, Switzerland, Germany, and the Czech Republic (Table 5). Based on the current knowledge of the geographical distribution of *Ca. N. mikurensis* in questing *I. ricinus* (Table 1), neehrlichiosis in (immune-compromised) patients are expected to occur basically all over Europe (Hansford et al. 2014; Jahfari et al. 2012). Less than half of these patients recalled tick exposure, making this a poor predictor of neehrlichiosis. Most patients had ongoing or recent immune suppressive treatment or were asplenic. The most frequent symptoms were fever, localized pain in muscles and/or joints, vascular and thromboembolic events (Grankvist et al. 2014). Typical laboratory findings were elevated C-reactive protein, leukocytosis with neutrophilia, and anemia.

Generally, patients recovered upon standard treatment with doxycycline. *Candidatus N. mikurensis* is apparently able to infect immune competent people as well; a recent study from China reported seven cases of *Ca. N. mikurensis* infection after screening blood samples from 622 febrile patients by PCR (1.1 %). All patients had a relatively mild disease: fever, headache, and malaise were reported for all seven patients, and none had a history of underlying immune-compromising conditions (Li et al. 2012). It was not reported whether the patients recovered upon treatment with antibiotics. Five cases of asymptomatic *Ca. N. mikurensis* infections in immune-competent individuals (1.6 %) were found by screening blood samples from 316 foresters in Poland by PCR (Welc-Faleciak et al. 2014b). How long *Ca. N. mikurensis* may circulate in healthy individuals is unknown. It is also unclear whether *Ca. N. mikurensis* infections may be transmitted via blood transfusion as described in eight cases of *A. phagocytophilum* infection (Townsend et al. 2014) and one case of ehrlichiosis (Regan et al. 2013). To what extent co-infections of *Ca. N. mikurensis* with other tick-borne pathogens, e.g. *Borrelia burgdorferi* s.l., can

**Table 5** Reported human cases of neohfrichiosis (until December 2014)

Year	Location	Case			Cause of immune suppression	References
		Gender	Age (years)			
2007	Germany	Male	69		Immunosuppressive therapy	von Loewenich et al. (2010)
2008	Germany	Male	57		Previously healthy	von Loewenich et al. (2010)
2009	Czech Republic	Female	55		Mantle cell lymphoma	Pekova et al. (2011)
	Sweden	Male	77		Chronic lymphocytic leukemia, asplenic	Welinder-Olsson et al. (2010)
2011	Switzerland	Male	61		Coronary artery bypass grafting	Fehr et al. (2010)
	Czech Republic	Male	58		Liver transplantation and splenectomy	Pekova et al. (2011)
	Switzerland	Male	68		Chronic lymphocytic leukemia	Maurer et al. (2013)
	Sweden	Male	77		B cell chronic lymphocytic leukemia, splenectomy	Grankvist et al. (2014)
2012	Switzerland	Female	67		Follicular lymphoma, (inborn) asplenic	Grankvist et al. (2014)
2013	Switzerland	Male	58		Follicular lymphoma	Maurer et al. (2013)
	Sweden	Female	67		T-cell large granular lymphoma splenectomy	Grankvist et al. (2014)
2014		Male	54		Psoriasis, immunosuppressive therapy	Grankvist et al. (2014)
	Sweden	Male	59		Rheumatoid arthritis, splenectomy	Grankvist et al. (2014)
		Female	71		Immunosuppressive therapy	Andréasson et al. (2015)

All but one patient had immune-compromised preconditions. The table is compiled from tables in Jalfari et al. (2012) and Grankvist et al. (2014)

affect disease severity has not been investigated so far (Horowitz et al. 2013; Lantos and Wormser 2014).

## ***Candidatus* N. mikurensis infection and clinical cases in domestic animals**

*Candidatus* N. mikurensis has been identified as one of the most prevalent microorganism in *I. ricinus*, indicating frequent exposure of animals to this potentially pathogenic species (Michelet et al. 2014). So far, one single case of the disease in an 8-year-old female dog has been described by Diniz et al. (2011). After ovariohysterectomy and mastectomy, the dog became lethargic and developed a profuse subcutaneous haemorrhage around the surgery site. Three days after surgery, the dog developed a subcutaneous hematoma. Laboratory findings included thrombocytopenia and mild anaemia, whereas serum chemistry profile and coagulation time showed normal results. Antibodies against vector-borne pathogens were not detected whereas PCR, performed 5 days after surgery, was positive for *Ehrlichia/Anaplasma* spp. Unfortunately, sequencing was not performed but treatment with doxycycline resulted in clinical stabilisation and resolution of the thrombocytopenia. Two months later, the dog was neutropenic, and this finding persisted over the next 2 months despite repeated treatments with doxycycline for 4 weeks. At that moment, *Ca. N. mikurensis* was confirmed by sequencing, and the treatment resulted in the elimination of the organism from the bloodstream. Haematological changes from the first and the second infection varied, and it was not possible to conclude whether *Ca. N. mikurensis* had caused thrombocytopenia, mild anaemia or neutropenia.

Based on the described findings in this dog, it is rather difficult to compare the clinical signs with the changes described in the human cases. Neutropenia persisted despite the elimination of *Ca. N. mikurensis* and deferred from the first findings. Based on the facts that a febrile stage was absent and there were non-specific haematological changes, it may be concluded that *Ca. N. mikurensis* was not the single cause of disease in this particular case. Recently, in a retrospective study with archival tissues from 19 dogs which died from haemolytic anaemia, *Ca. N. mikurensis* was detected and confirmed by sequencing in a 3.5 months old puppy (Beck et al. 2014a). This finding represents the first case in which *Ca. N. mikurensis* may have been pathogenic for younger dogs. There is no evidence of pathogenicity in other domestic animal species. A big progress in the knowledge of several canine vector-borne diseases has been made with frequent use of fast tests in small animal clinics. Such a test for detecting infection with *Ca. N. mikurensis* does not exist so far. Therefore, the role of *Ca. N. mikurensis* in disease development in companion animals remains unclear.

## **Public health relevance**

Based on current findings, incidences and estimations of the disease burden caused by *Ca. N. mikurensis* infections cannot be given—they will certainly be much lower than for Lyme borreliosis, tick-borne encephalitis, or Mediterranean spotted fever, but higher than for human granulocytic anaplasmosis in Europe, with the ubiquitous prevalence of the causing agent in ticks but almost no reported human cases. The public health relevance of *Ca. N. mikurensis* is currently not caused by high mortality or morbidity rates in the general population, but probably rather by the obliviousness of most health professionals

for neehrlichiosis (Grankvist et al. 2014; Maurer et al. 2013). Medical specialists would recognize the atypical symptoms of patients with neehrlichiosis, but are currently unable to provide an etiological diagnosis. As a consequence, these patients may not get an adequate antibiotic treatment, or only very late, or they may get ‘treatment’ based on a different diagnosis, e.g. Lyme borreliosis. Increasing awareness (alone) may not be sufficient. The development of clinical case definitions for ‘probable’ and ‘possible’ neehrlichiosis, together with laboratory diagnostics, will be highly supportive for medical specialists in order to gain ‘confirmed’ cases and thus counts necessary to judge the impact of this disease on public health.

In The Netherlands alone, it can be estimated that approximately 60,000 persons are annually bitten by ticks infected with *Ca. N. mikurensis*, compared to 183,000 that are bitten by ticks infected with *B. burgdorferi* sensu lato, the causative agent of Lyme borreliosis (Fonville et al. 2014; Jahfari et al. 2012). Thus, exposure of people to ticks infected with *Ca. N. mikurensis* is relatively high, and is expected to be on the rise. In The Netherlands, the incidence of (reported) tick bites and erythema migrans has increased three- to fourfold over the past 2 decades, which could be explained partially by a concomitant increase in diagnostic requests and/or the total number of questing *I. ricinus* (Hofhuis et al. 2015; Sprong et al. 2012). Environmental (e.g. landscape management and climate change), socio-economic and demographic factors (e.g. population aging and life-styles) synergistically increase the risk of acquiring tick-borne diseases (Godfrey and Randolph 2011; Medlock et al. 2013; Sprong et al. 2012; Stefanoff et al. 2012). Furthermore, an ageing population and advanced treatments for patients with haematological, oncologic and rheumatological diseases increase the numbers of immune-compromised persons in the general population who have a higher risk for developing neehrlichiosis (Cascio et al. 2011).

## Gaps of knowledge and need for future research

Until now, *Ca. N. mikurensis* has not been cultured. Its closest relative, *Ca. Neehrlichia lotoris*, is cultivatable in a tick cell line and this might be a promising approach for *Ca. N. mikurensis* too (Yabsley et al. 2008a). A culture is still the key to the development of molecular and serological diagnostics tools, as well as tests of the specific sensitivity to antibiotics of this bacterium (Raoult 2014). These gaps in knowledge indicate the necessity of research for revealing the impact on human and animal health. The majority of published research deals with the ecology of *Ca. N. mikurensis* and detection in human patients. Further research should also focus on determining the role of *Ca. N. mikurensis* in clinical cases in animals. In addition, it is necessary to determine the role of *Ca. N. mikurensis* in disease development with concurrent infections. To determine the full genome sequence would be widely beneficial: it could be used to devise specific molecular diagnostic tests and it could lead to the identification of protein-coding regions that might be used as recombinant proteins to develop serological diagnostics in case the attempts to isolate *Ca. N. mikurensis* in vitro continue to fail. The genomic information may likewise be useful to identify regions that are involved in pathogenicity during host cell infection. Finally, genomic regions with considerable levels of heterogeneity may be identified, which in turn may allow strain differentiation, phylogenetic analysis or identification of particular transmission cycles.

Although a considerable amount of field data has been gathered over the last decade, no systematic study solely on *Ca. N. mikurensis* has been performed, neither in a natural

environment nor using different rodent species in the laboratory, in order to follow the time course of infection on the individual and the population level. These efforts would help determine the real threat that *Ca. N. mikurensis* poses to human and animal health, develop serological methods, and investigate genetic diversity focusing on isolates that cause clinical signs in humans.

## Conclusion

*Candidatus N. mikurensis* is one of those examples where the pathogen was detected before the disease it causes. This course of events is expected to be rather the rule than the exception in the future (Tijssen-Klasen et al. 2014). Several important aspects of *Ca. N. mikurensis* have recently been assessed: rodents are its vertebrate reservoir hosts, it is widely distributed in the vector tick species *I. ricinus* and *I. persulcatus* and it may cause severe diseases in immune-compromised patients. Above all, the isolation of *Ca. N. mikurensis* should be attempted in order to develop diagnostic tools and to experimentally investigate its transmission cycle and pathogenic properties in controlled laboratory experiments.

**Acknowledgments** This work was done under the frame of WG5 of COST action TD1303 EurNegVec. We thank Alexander Mathis for some suggestions on the manuscript.

**Conflict of interest** The authors declare that they have no conflict of interest.

## References

- Alekseev AN, Dubinina HV, Van De Pol I, Schouls LM (2001) Identification of *Ehrlichia* spp. and *Borrelia burgdorferi* in *Ixodes* ticks in the baltic regions of Russia. *J Clin Microbiol* 39:2237–2242. doi:10.1128/JCM.39.6.2237-2242.2001
- Andersson M, Raberg L (2011) Wild rodents and novel human pathogen *Candidatus Neoehrlichia mikurensis*, Southern Sweden. *Emerg Infect Dis* 17:1716–1718. doi:10.3201/eid1709.101058
- Andersson M, Scherman K, Raberg L (2014a) Infection dynamics of the tick-borne pathogen ‘*Candidatus Neoehrlichia mikurensis*’ and coinfections with *Borrelia afzelii* in bank voles in Southern Sweden. *Appl Environ Microbiol* 80:1645–1649. doi:10.1128/AEM.03469-13
- Andersson M, Zaghdoudi-Allan N, Tamba P, Stefanache M, Chitimia L (2014b) Co-infection with ‘*Candidatus Neoehrlichia mikurensis*’ and *Borrelia afzelii* in an *Ixodes ricinus* tick that has bitten a human in Romania. *Ticks Tick Borne Dis* 5:706–708. doi:10.1016/j.ttbdis.2014.05.013
- Andréasson K, Jönsson G, Lindell P, Gülfe A, Ingvarsson R, Lindqvist E, Saxne T, Grankvist A, Wennerås C, Marsal J (2015) Recurrent fever caused by *Candidatus Neoehrlichia mikurensis* in a rheumatoid arthritis patient treated with rituximab. *Rheumatology (Oxford)* 54:369–371. doi:10.1093/rheumatology/keu441
- Aubry P, Geale DW (2011) A review of bovine anaplasmosis. *Transbound Emerg Dis* 58:1–30. doi:10.1111/j.1865-1682.2010.01173.x
- Beck A, Huber D, Antolić M, Anzulović Ž, Reil I, Beck, R (2014a). Molecular retrospective study of canine infectious haemolytic anaemias. *Cutting Edge Pathology*, 2nd Joint European Congress of the ESVP, ESTP and ECVF Berlin pp. 84–84
- Beck R, Čubrčić Čurik V, Račić I, Šprem N, Vujnović A (2014b) Identification of *Candidatus Neoehrlichia mikurensis* and *Anaplasma* species in wildlife from Croatia. *Proceedings of the 1st Conference on Neglected Vectors and Vector-Borne Diseases (EurNegVec) Cluj-Napoca Parasites & Vectors*, 7(Suppl 1): O28
- Beninati T, Piccolo G, Rizzoli A, Genchi C, Bandi C (2006) *Anaplasmataceae* in wild rodents and roe deer from Trento Province (northern Italy). *Eur J Clin Microbiol Infect Dis* 25:677–678

- Bowman DD (2011) Introduction to the alpha-proteobacteria: *Wolbachia* and *Bartonella*, *Rickettsia*, *Bruceella*, *Ehrlichia*, and *Anaplasma*. Top Companion Anim Med 26:173–177. doi:10.1053/j.tcam.2011.09.002
- Burri C, Schumann O, Schumann C, Gern L (2014) Are *Apodemus* spp. mice and *Myodes glareolus* reservoirs for *Borrelia miyamotoi*, *Candidatus* Neohrlichia mikurensis, *Rickettsia helvetica*, *R. monacensis* and *Anaplasma phagocytophilum*? Ticks Tick Borne Dis 5:245–251. doi:10.1016/j.ttbdis.2013.11.007
- Capelli G, Ravagnan S, Montarsi F, Ciocchetta S, Cazzin S, Porcellato E, Babiker AM, Cassini R, Salviato A, Cattoli G, Otranto D (2012) Occurrence and identification of risk areas of *Ixodes ricinus*-borne pathogens: a cost-effectiveness analysis in north-eastern Italy. Parasit Vectors 5:61. doi:10.1186/1756-3305-5-61
- Cascio A, Bosilkovski M, Rodriguez-Morales AJ, Pappas G (2011) The socio-ecology of zoonotic infections. Clin Microbiol Infect 17:336–342. doi:10.1111/j.1469-0691.2010.03451.x
- Coipan EC, Jahfari S, Fonville M, Maassen CB, van der Giessen J, Takken W, Takumi K, Sprong H (2013) Spatiotemporal dynamics of emerging pathogens in questing *Ixodes ricinus*. Front Cell Infect Microbiol 3:36. doi:10.3389/fcimb.2013.00036
- Derdáková M, Václav R, Pangrácova-Blaňárová L, Selyemová D, Koči J, Walder G, Špitalská E (2014) *Candidatus* Neohrlichia mikurensis and its co-circulation with *Anaplasma phagocytophilum* in *Ixodes ricinus* ticks across ecologically different habitats of Central Europe. Parasit Vectors 7:160. doi:10.1186/1756-3305-7-160
- Diniz PP, Schulz BS, Hartmann K, Breitschwerdt EB (2011) '*Candidatus* Neohrlichia mikurensis' infection in a dog from Germany. J Clin Microbiol 49:2059–2062. doi:10.1128/JCM.02327-10
- Dumler JS, Barbet AF, Bekker CP, Dasch GA, Palmer GH, Ray SC, Rikihisa Y, Rurangirwa FR (2001) Reorganization of genera in the families Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: unification of some species of *Ehrlichia* with *Anaplasma*, *Cowdria* with *Ehrlichia* and *Ehrlichia* with *Neorickettsia*, descriptions of six new species combinations and designation of *Ehrlichia equi* and 'HGE agent' as subjective synonyms of *Ehrlichia phagocytophila*. Int J Syst Evol Microbiol 51:2145–2165
- Dumler JS, Madigan JE, Pusterla N, Bakken JS (2007) Ehrlichioses in humans: epidemiology, clinical presentation, diagnosis, and treatment. Clin Infect Dis 45(Suppl 1):S45–S51. doi:10.1086/518146
- Fehr JS, Bloemberg GV, Ritter C, Hombach M, Luscher TF, Weber R, Keller PM (2010) Septicemia caused by tick-borne bacterial pathogen *Candidatus* Neohrlichia mikurensis. Emerg Infect Dis 16:1127–1129. doi:10.3201/eid1607.091907
- Fertner ME, Molbak L, Boye Pihl TP, Fomsgaard A, Bodker R (2012) First detection of tick-borne '*Candidatus* Neohrlichia mikurensis' in Denmark 2011. Euro Surveill 17:20096
- Földvári G, Jahfari S, Rigó K, Jablonszky M, Szekeres S, Majoros G, Tóth M, Molnár V, Coipan EC, Sprong H (2014) *Candidatus* Neohrlichia mikurensis and *Anaplasma phagocytophilum* in urban hedgehogs. Emerg Infect Dis 20:496–498. doi:10.3201/eid2003.130935
- Fonville M, Friesema IH, Hengeveld PD, Docters van Leeuwen A, Jahfari S, Harms MG, van Vliet AJ, Hofhuis A, van Pelt W, Sprong H, van den Wijngaard CC (2014) Human exposure to tickborne relapsing fever spirochete *Borrelia miyamotoi*, the Netherlands. Emerg Infect Dis 20:1244–1245. doi:10.3201/eid2007.131525
- Friedman CS, Andree KB, Beauchamp KA, Moore JD, Robbins TT, Shields JD, Hedrick RP (2000) '*Candidatus* Xenohaliotis californiensis', a newly described pathogen of abalone, *Haliotis* spp., along the west coast of North America. Int J Syst Evol Microbiol 50(Pt 2):847–855
- Glatz M, Mullegger RR, Maurer F, Fingerle V, Achermann Y, Wilske B, Bloemberg GV (2014) Detection of *Candidatus* Neohrlichia mikurensis, *Borrelia burgdorferi* sensu lato genospecies and *Anaplasma phagocytophilum* in a tick population from Austria. Ticks Tick Borne Dis 5:139–144. doi:10.1016/j.ttbdis.2013.10.006
- Godfrey ER, Randolph SE (2011) Economic downturn results in tick-borne disease upsurge. Parasit Vectors 4:35. doi:10.1186/1756-3305-4-35
- Grankvist A, Andersson PO, Mattsson B, Sender M, Vaht K, Höper L, Sakiniene E, Trysberg E, Stenson M, Fehr J, Pekova S, Bogdan C, Bloemberg G, Wennerås C (2014) Infections with the tick-borne bacterium '*Candidatus* Neohrlichia mikurensis' mimic noninfectious conditions in patients with B cell malignancies or autoimmune diseases. Clin Infect Dis 58:1716–1722. doi:10.1093/cid/ciu189
- Hansford KM, Fonville M, Jahfari S, Sprong H, Medlock JM (2014). *Borrelia miyamotoi* in host-seeking *Ixodes ricinus* ticks in England Epidemiol Infect:1–9 doi:10.1017/S0950268814001691
- Heyman P, Cochez C, Hofhuis A, van der Giessen J, Sprong H, Porter SR, Losson B, Saegerman C, Donoso-Mantke O, Niedrig M, Papa A (2010) A clear and present danger: tick-borne diseases in Europe. Expert Rev Anti Infect Ther 8:33–50. doi:10.1586/eri.09.118

- Hofhuis A, Harms M, van den Wijngaard C, Sprong H, van Pelt W (2015) Continuing increase of tick bites and Lyme disease between 1994 and 2009. *Ticks Tick Borne Dis* 6:69–74. doi:[10.1016/j.ttbdis.2014.09.006](https://doi.org/10.1016/j.ttbdis.2014.09.006)
- Hornok S, Meli ML, Gonczi E, Hofmann-Lehmann R (2013) First evidence of *Candidatus* Neoehrlichia mikurensis in Hungary. *Parasit Vectors* 6:267. doi:[10.1186/1756-3305-6-267](https://doi.org/10.1186/1756-3305-6-267)
- Hornok S, Kováts D, Csörgő T, Meli ML, Gónczi E, Hadnagy Z, Takács N, Farkas R, Hofmann-Lehmann R (2014) Birds as potential reservoirs of tick-borne pathogens: first evidence of bacteraemia with *Rickettsia helvetica*. *Parasit Vectors* 7:128. doi:[10.1186/1756-3305-7-128](https://doi.org/10.1186/1756-3305-7-128)
- Horowitz HW, Aguero-Rosenfeld ME, Holmgren D, McKenna D, Schwartz I, Cox ME, Wormser GP (2013) Lyme disease and human granulocytic anaplasmosis coinfection: impact of case definition on coinfection rates and illness severity. *Clin Infect Dis* 56:93–99. doi:[10.1093/cid/cis852](https://doi.org/10.1093/cid/cis852)
- Jahfari S, Fonville M, Hengeveld P, Reusken C, Scholte EJ, Takken W, Heyman P, Medlock JM, Heylen D, Kleve J, Sprong H (2012) Prevalence of *Neoehrlichia mikurensis* in ticks and rodents from North–west Europe. *Parasit Vectors* 5:74. doi:[10.1186/1756-3305-5-74](https://doi.org/10.1186/1756-3305-5-74)
- Kamani J, Baneth G, Mumcuoglu KY, Waziri NE, Eyal O, Guthmann Y, Harrus S (2013) Molecular detection and characterization of tick-borne pathogens in dogs and ticks from Nigeria. *PLoS Negl Trop Dis* 7:e2108. doi:[10.1371/journal.pntd.0002108](https://doi.org/10.1371/journal.pntd.0002108)
- Kawahara M, Rikihisa Y, Isogai E, Takahashi M, Misumi H, Suto C, Shibata S, Zhang C, Tsuji M (2004) Ultrastructure and phylogenetic analysis of ‘*Candidatus* Neoehrlichia mikurensis’ in the family Anaplasmataceae, isolated from wild rats and found in *Ixodes ovatus* ticks. *Int J Syst Evol Microbiol* 54:1837–1843. doi:[10.1099/ijs.0.63260-0](https://doi.org/10.1099/ijs.0.63260-0)
- Koutouris M, Santos AS, Dumler JS, Brouqui P (2005) Distribution of ‘*Ehrlichia walkeri*’ in *Ixodes ricinus* (Acari: Ixodidae) from the northern part of Italy. *J Med Entomol* 42:82–85
- Krücken J, Schreiber C, Maaz D, Kohn M, Demeler J, Beck S, Schein E, Olias P, Richter D, Matuschka FR, Pachnicke S, Krieger K, Kohn B, von Samson-Himmelstjerna G (2013) A novel high-resolution melt PCR assay discriminates *Anaplasma phagocytophilum* and ‘*Candidatus* Neoehrlichia mikurensis’. *J Clin Microbiol* 51:1958–1961. doi:[10.1128/JCM.00284-13](https://doi.org/10.1128/JCM.00284-13)
- Lantos PM, Wormser GP (2014) Chronic coinfections in patients diagnosed with chronic lyme disease: a systematic review. *Am J Med* 127:1105–1110. doi:[10.1016/j.amjmed.2014.05.036](https://doi.org/10.1016/j.amjmed.2014.05.036)
- Li H, Jiang JF, Liu W, Zheng YC, Huo QB, Tang K, Zuo SY, Liu K, Jiang BG, Yang H, Cao WC (2012) Human infection with *Candidatus* Neoehrlichia mikurensis, China. *Emerg Infect Dis* 18:1636–1639. doi:[10.3201/eid1810.120594](https://doi.org/10.3201/eid1810.120594)
- Li H, Jiang J, Tang F, Sun Y, Li Z, Zhang W, Gong Z, Liu K, Yang H, Liu W, Cao W (2013) Wide distribution and genetic diversity of ‘*Candidatus* Neoehrlichia mikurensis’ in rodents from China. *Appl Environ Microbiol* 79:1024–1027. doi:[10.1128/AEM.02917-12](https://doi.org/10.1128/AEM.02917-12)
- Lommano E, Bertaiola L, Dupasquier C, Gern L (2012) Infections and coinfections of questing *Ixodes ricinus* ticks by emerging zoonotic pathogens in Western Switzerland. *Appl Environ Microbiol* 78:4606–4612. doi:[10.1128/AEM.07961-11](https://doi.org/10.1128/AEM.07961-11)
- Lommano E, Dvorak C, Vallotton L, Jenni L, Gern L (2014) Tick-borne pathogens in ticks collected from breeding and migratory birds in Switzerland. *Ticks Tick Borne Dis* 5:871–882. doi:[10.1016/j.ttbdis.2014.07.001](https://doi.org/10.1016/j.ttbdis.2014.07.001)
- Maurer FP, Keller PM, Beuret C, Joha C, Achermann Y, Gubler J, Bircher D, Karrer U, Fehr J, Zimmerli L, Bloemberg GV (2013) Close geographic association of human neoehrlichiosis and tick populations carrying ‘*Candidatus* Neoehrlichia mikurensis’ in eastern Switzerland. *J Clin Microbiol* 51:169–176. doi:[10.1128/JCM.01955-12](https://doi.org/10.1128/JCM.01955-12)
- Medlock JM, Hansford KM, Bormane A, Derdakova M, Estrada-Peña A, George JC, Golovljova I, Jaenson TG, Jensen JK, Jensen PM, Kazimirova M, Oteo JA, Papa A, Pfister K, Plantard O, Randolph SE, Rizzoli A, Santos-Silva MM, Sprong H, Vial L, Hendrickx G, Zeller H, Van Bortel W (2013) Driving forces for changes in geographical distribution of *Ixodes ricinus* ticks in Europe. *Parasit Vectors* 6:1. doi:[10.1186/1756-3305-6-1](https://doi.org/10.1186/1756-3305-6-1)
- Michelet L, Delannoy S, Devillers E, Umhang G, Aspan A, Juremalm M, Chirico J, van der Wal FJ, Sprong H, Boye Pihl TP, Klitgaard K, Bødker R, Fach P, Moutailler S (2014) High-throughput screening of tick-borne pathogens in Europe. *Front Cell Infect Microbiol* 4:103. doi:[10.3389/fcimb.2014.00103](https://doi.org/10.3389/fcimb.2014.00103)
- Movila A, Alekseev AN, Dubinina HV, Toderas I (2013a) Detection of tick-borne pathogens in ticks from migratory birds in the Baltic region of Russia. *Med Vet Entomol* 27:113–117. doi:[10.1111/j.1365-2915.2012.01037.x](https://doi.org/10.1111/j.1365-2915.2012.01037.x)
- Movila A, Toderas I, Uspenskaia I, Conovalov J (2013b) Molecular detection of tick-borne pathogens in *Ixodes ricinus* from Moldova collected in 1960. *Ticks Tick Borne Dis* 4:359–361. doi:[10.1016/j.ttbdis.2012.12.004](https://doi.org/10.1016/j.ttbdis.2012.12.004)

- Munderloh UG, Yabsley MJ, Murphy SM, Luttrell MP, Howerth EW (2007) Isolation and establishment of the raccoon *Ehrlichia*-like agent in tick cell culture. *Vector Borne Zoonotic Dis* 7:418–425. doi:10.1089/vbz.2007.0640
- Naitou H, Kawaguchi D, Nishimura Y, Inayoshi M, Kawamori F, Masuzawa T, Hiroi M, Kurashige H, Kawabata H, Fujita H, Ohashi N (2006) Molecular identification of *Ehrlichia* species and ‘*Candidatus* Neoehrlichia mikurensis’ from ticks and wild rodents in Shizuoka and Nagano Prefectures, Japan. *Microbiol Immunol* 50:45–51
- Obiegala A, Pfeffer M, Pfister K, Tiedemann T, Thiel C, Balling A, Karnath C, Woll D, Silaghi C (2014) *Candidatus* Neoehrlichia mikurensis and *Anaplasma phagocytophilum*: prevalences and investigations on a new transmission path in small mammals and ixodid ticks. *Parasit Vectors* 7:563. doi:10.1186/s13071-014-0563-x
- Otranto D, Dantas-Torres F, Giannelli A, Latrofa MS, Cascio A, Cazzin S, Ravagnan S, Montarsi F, Zanzani SA, Manfredi MT, Capelli G (2014) Ticks infesting humans in Italy and associated pathogens. *Parasit Vectors* 7:328. doi:10.1186/1756-3305-7-328
- Palomar AM, García-Alvarez L, Santibanez S, Portillo A, Oteo JA (2014) Detection of tick-borne ‘*Candidatus* Neoehrlichia mikurensis’ and *Anaplasma phagocytophilum* in Spain in 2013. *Parasit Vectors* 7:57. doi:10.1186/1756-3305-7-57
- Pan H, Liu S, Ma Y, Tong S, Sun Y (2003) *Ehrlichia*-like organism gene found in small mammals in the suburban district of Guangzhou of China. *Ann N Y Acad Sci* 990:107–111
- Pangráčová L, Derdákóvá M, Pekárik L, Hviščová I, Víchová B, Stanko M, Hlavatá H, Peňko B (2013) Ixodes ricinus abundance and its infection with the tick-borne pathogens in urban and suburban areas of Eastern Slovakia. *Parasit Vectors* 6:238. doi:10.1186/1756-3305-6-238
- Pekova S, Vydra J, Kabickova H, Frankova S, Haugvicova R, Mazal O, Cmejla R, Hardekopf DW, Jancuskova T, Kozak T (2011) *Candidatus* Neoehrlichia mikurensis infection identified in 2 hematologic patients: benefit of molecular techniques for rare pathogen detection. *Diagn Microbiol Infect Dis* 69:266–270. doi:10.1016/j.diagmicrobio.2010.10.004
- Raoult D (2014) Uncultured *Candidatus* Neoehrlichia mikurensis. *Clin Infect Dis* 59:1042. doi:10.1093/cid/ciu491
- Rar VA, Livanova NN, Panov VV, Doroschenko EK, Pukhovskaya NM, Vysochina NP, Ivanov LI (2010) Genetic diversity of *Anaplasma* and *Ehrlichia* in the Asian part of Russia. *Ticks Tick Borne Dis* 1:57–65. doi:10.1016/j.tbd.2010.01.002
- Regan J, Matthias J, Green-Murphy A, Stanek D, Bertholf M, Pritt BS, Sloan LM, Kelly AJ, Singleton J, McQuiston JH, Hovevar SN, Whittle JP (2013) A confirmed *Ehrlichia ewingii* infection likely acquired through platelet transfusion. *Clin Infect Dis* 56:e105–e107. doi:10.1093/cid/cit177
- Richter D, Matuschka FR (2012) ‘*Candidatus* Neoehrlichia mikurensis’, *Anaplasma phagocytophilum*, and Lyme disease spirochetes in questing European vector ticks and in feeding ticks removed from people. *J Clin Microbiol* 50:943–947. doi:10.1128/JCM.05802-11
- Richter D, Kohn C, Matuschka FR (2013) Absence of *Borrelia* spp., *Candidatus* Neoehrlichia mikurensis, and *Anaplasma phagocytophilum* in questing adult *Dermacentor reticulatus* ticks. *Parasitol Res* 112:107–111. doi:10.1007/s00436-012-3110-8
- Schouls LM, Van De Pol I, Rijpkema SG, Schot CS (1999) Detection and identification of *Ehrlichia*, *Borrelia burgdorferi* sensu lato, and *Bartonella* species in Dutch *Ixodes ricinus* ticks. *J Clin Microbiol* 37:2215–2222
- Schreiber C, Krücken J, Beck S, Maaz D, Pachnicke S, Krieger K, Gross M, Kohn B, von Samson-Himmelstjerna G (2014) Pathogens in ticks collected from dogs in Berlin/Brandenburg, Germany. *Parasit Vectors* 7:535. doi:10.1186/s13071-014-0535-1
- Shpynov S, Fournier PE, Rudakov N, Tarasevich I, Raoult D (2006) Detection of members of the genera *Rickettsia*, *Anaplasma*, and *Ehrlichia* in ticks collected in the Asiatic part of Russia. *Ann N Y Acad Sci* 1078:378–383. doi:10.1196/annals.1374.075
- Silaghi C, Woll D, Mahling M, Pfister K, Pfeffer M (2012) *Candidatus* Neoehrlichia mikurensis in rodents in an area with sympatric existence of the hard ticks *Ixodes ricinus* and *Dermacentor reticulatus*, Germany. *Parasit Vectors* 5:285. doi:10.1186/1756-3305-5-285
- Silaghi C, Pfister K, Overzier E (2014) Molecular investigation for bacterial and protozoan tick-borne pathogens in wild boars (*Sus scrofa*) from southern Germany. *Vector Borne Zoonotic Dis* 14:371–373. doi:10.1089/vbz.2013.1495
- Špitálská E, Boldis V, Kostanová Z, Kocianová E, Stefanidesová K (2008) Incidence of various tick-borne microorganisms in rodents and ticks of central Slovakia. *Acta Virol* 52:175–179
- Sprong H, Hofhuis A, Gassner F, Takken W, Jacobs F, van Vliet AJ, van Ballegooijen M, van der Giessen J, Takumi K (2012) Circumstantial evidence for an increase in the total number and activity of *Borrelia*-infected *Ixodes ricinus* in the Netherlands. *Parasit Vectors* 5:294. doi:10.1186/1756-3305-5-294

- Stefanoff P, Rosinska M, Samuels S, White DJ, Morse DL, Randolph SE (2012) A national case-control study identifies human socio-economic status and activities as risk factors for tick-borne encephalitis in Poland. *PLoS ONE* 7:e45511. doi:[10.1371/journal.pone.0045511](https://doi.org/10.1371/journal.pone.0045511)
- Szekeress S, Claudia Coipan E, Rigo K, Majoros G, Jahfari S, Sprong H, Földvári G (2015) *Candidatus Neoehrlichia mikurensis* and *Anaplasma phagocytophilum* in natural rodent and tick communities in Southern Hungary. *Ticks Tick Borne Dis* 6:111–116. doi:[10.1016/j.ttbdis.2014.10.004](https://doi.org/10.1016/j.ttbdis.2014.10.004)
- Tabara K, Arai S, Kawabuchi T, Itagaki A, Ishihara C, Satoh H, Okabe N, Tsuji M (2007) Molecular survey of *Babesia microti*, *Ehrlichia* species and *Candidatus neoehrlichia mikurensis* in wild rodents from Shimane Prefecture, Japan. *Microbiol Immunol* 51:359–367
- Tijssse-Klasen E, Koopmans MP, Sprong H (2014) Tick-borne pathogen—reversed and conventional discovery of disease. *Front Public Health*. 2:73. doi:[10.3389/fpubh.2014.00073](https://doi.org/10.3389/fpubh.2014.00073)
- Townsend RL, Moritz ED, Fialkow LB, Berardi V, Stramer SL (2014) Probable transfusion-transmission of *Anaplasma phagocytophilum* by leukoreduced platelets. *Transfusion* 54:2828–2832. doi:[10.1111/trf.12675](https://doi.org/10.1111/trf.12675)
- Vayssier-Taussat M, Le Rhun D, Buffet JP, Maaoui N, Galan M, Guivier E, Charbonnel N, Cosson JF (2012) *Candidatus Neoehrlichia mikurensis* in bank voles, France. *Emerg Infect Dis* 18:2063–2065. doi:[10.3201/eid1812.120846](https://doi.org/10.3201/eid1812.120846)
- Venclikova K, Rudolf I, Mendel J, Betasova L, Hubalek Z (2014) Rickettsiae in questing *Ixodes ricinus* ticks in the Czech Republic. *Ticks Tick Borne Dis* 5:135–138. doi:[10.1016/j.ttbdis.2013.09.008](https://doi.org/10.1016/j.ttbdis.2013.09.008)
- Vichova B et al (2014) *Anaplasma* infections in ticks and reservoir host from Slovakia. *Infect Genet Evol* 22:265–272. doi:[10.1016/j.meegid.2013.06.003](https://doi.org/10.1016/j.meegid.2013.06.003)
- Von Loewenich FD, Stumpf G, Baumgarten BU, Rollinghoff M, Dumler JS, Bogdan C (2003) Human granulocytic ehrlichiosis in Germany: evidence from serological studies, tick analyses, and a case of equine ehrlichiosis. *Ann N Y Acad Sci* 990:116–117
- Von Loewenich FD, Geissdorfer W, Disque C, Matten J, Schett G, Sakka SG, Bogdan C (2010) Detection of ‘*Candidatus Neoehrlichia mikurensis*’ in two patients with severe febrile illnesses: evidence for a European sequence variant. *J Clin Microbiol* 48:2630–2635. doi:[10.1128/JCM.00588-10](https://doi.org/10.1128/JCM.00588-10)
- Welc-Faleciak R, Kowalec M, Karbowiak G, Bajer A, Behnke JM, Sinski E (2014a) Rickettsiaceae and Anaplasmataceae infections in *Ixodes ricinus* ticks from urban and natural forested areas of Poland. *Parasit Vectors* 7:121. doi:[10.1186/1756-3305-7-121](https://doi.org/10.1186/1756-3305-7-121)
- Welc-Faleciak R, Sinski E, Kowalec M, Zajkowska J, Pancewicz SA (2014b) Asymptomatic ‘*Candidatus Neoehrlichia mikurensis*’ infections in immunocompetent humans. *J Clin Microbiol* 52:3072–3074. doi:[10.1128/JCM.00741-14](https://doi.org/10.1128/JCM.00741-14)
- Welinder-Olsson C, Kjellin E, Vaht K, Jacobsson S, Wenneras C (2010) First case of human ‘*Candidatus Neoehrlichia mikurensis*’ infection in a febrile patient with chronic lymphocytic leukemia. *J Clin Microbiol* 48:1956–1959. doi:[10.1128/JCM.02423-09](https://doi.org/10.1128/JCM.02423-09)
- Yabsley MJ, Murphy SM, Luttrell MP, Wilcox BR, Howerth EW, Munderloh UG (2008a) Characterization of ‘*Candidatus Neoehrlichia lotoris*’ (family Anaplasmataceae) from raccoons (*Procyon lotor*). *Int J Syst Evol Microbiol* 58:2794–2798. doi:[10.1099/ijs.0.65836-0](https://doi.org/10.1099/ijs.0.65836-0)
- Yabsley MJ, Murphy SM, Luttrell MP, Wilcox BR, Ruckdeschel C (2008b) Raccoons (*Procyon lotor*), but not rodents, are natural and experimental hosts for an ehrlichial organism related to ‘*Candidatus Neoehrlichia mikurensis*’. *Vet Microbiol* 131:301–308. doi:[10.1016/j.vetmic.2008.04.004](https://doi.org/10.1016/j.vetmic.2008.04.004)